



Rat Salivary Gland Fibroblasts (RSGF) Catalog #R2670

Cell Specification

The salivary gland produces enzyme-containing saliva to lubricate the oral cavity and assist digestion. It consists of three pairs of major salivary glands and many minor ones in the oral cavity [1]. Histology studies show that the salivary gland is a compound tubuloalveolar gland, surrounded by connective tissue to further divide the gland into lobes and lobules. The secretory endpieces consist of a spherical mass of cells called acinus. Rat salivary gland fibroblasts (RSGF) are embryonic mesoderm derived mesenchymal cells. They are responsible for much of the synthesis of extracellular matrix including type I and/or type III collagen [2]. RSGF are very versatile and would provide a useful tool in studying collagenous fibroma and oral neoplasms [3].

RSGF from ScienCell Research Laboratories are isolated from rat salivary gland tissues. RSGF are cryopreserved at P1 and delivered frozen. Each vial contains $>5 \times 10^5$ cells in 1 ml volume. RSGF are characterized by immunofluorescence with antibodies specific to fibronectin. RSGF are negative for mycoplasma, bacteria, yeast and fungi. RSGF are guaranteed to further culture in the conditions provided by ScienCell Research Laboratories.

Recommended Medium

It is recommended to use Fibroblast Medium (FM, Cat. #2301) for the culturing of RSGF *in vitro*.

Product Use

RSGF are for research use only. They are not approved for human or animal use, or for application in *in vitro* diagnostic procedures.

Storage

Transfer cells directly and immediately from dry ice to liquid nitrogen upon receiving and keep the cells in liquid nitrogen until cell culture is needed for experiments.

Shipping

Dry ice.

References

- [1] Baker OJ. (2010) "Tight junctions in salivary epithelium." *J Biomed Biotechnol.* 2010: 278948.
- [2] Gabbiani G, Rungger-Brandle E. (1981) "The fibroblast." In Glynn LE, *Handbook of Inflammation, Vol. 3: Tissue Repair and Regeneration* (pp 1-50). Amsterdam: Elsevier.
- [3] Ide F, Shimoyama T, Horie N, Tanaka H. (1999) "Collagenous fibroma (desmoplastic fibroblastoma) presenting as a parotid mass." *J Oral Pathol Med.* 28: 465-8.

Instruction for culturing cells

Caution: Cryopreserved cells are very delicate. Thaw the vial in a 37°C water bath and return the cells to culture as quickly as possible with minimal handling!

Initiating the culture:

1. Prepare a poly-L-lysine coated culture vessel (2 $\mu\text{g}/\text{cm}^2$, T-75 flask is recommended). Add 10 ml of sterile water to a T-75 flask and then add 15 μl of poly-L-lysine stock solution (10 mg/ml, Cat. #0413). Leave the vessel in incubator overnight (minimum one hour at 37°C incubator).
2. Prepare complete medium. Decontaminate the external surfaces of medium bottle and medium supplement tubes with 70% ethanol and transfer them to a sterile field. Aseptically transfer supplement to the basal medium with a pipette. Rinse the tube with medium to recover the entire volume.
3. Rinse the poly-L-lysine coated vessel with sterile water twice and then add 15 ml of complete medium. Leave the vessel in the sterile field and proceed to thaw the cryopreserved cells.
4. Place the frozen vial in a 37°C water bath. Hold and rotate the vial gently until the contents completely thaw. Remove the vial from the water bath promptly, wipe it down with 70% ethanol and transfer it to the sterile field.
5. Remove the cap carefully without touching the interior threads. Gently resuspend and dispense the contents of the vial into the equilibrated, poly-L-lysine coated culture vessel. A seeding density of 8,000 cells/cm² is recommended.
Note: Dilution and centrifugation of cells after thawing are not recommended since these actions are more harmful to the cells than the effect of residual DMSO in the culture. It is also important that cells are plated in poly-L-lysine coated culture vessels to promote cell attachment.
6. Replace the cap or lid, and gently rock the vessel to distribute the cells evenly. Loosen cap if necessary to allow gas exchange.
7. Return the culture vessel to the incubator.
8. For the best result, do not disturb the culture for at least 16 hours after the culture has been initiated. Refresh culture medium the next day to remove the residual DMSO and unattached cells, then every other day thereafter.

Maintaining the culture:

1. Refresh supplemented culture medium the next morning after establishing a culture from cryopreserved cells.
2. Change the medium every three days thereafter, until the culture is approximately 70% confluent.
3. Once the culture reaches 70% confluence, change medium every other day until the culture is approximately 90% confluent.

Subculturing:

1. Subculture when the culture reaches 90% confluency or above.
2. Prepare poly-L-lysine coated culture vessels ($2 \mu\text{g}/\text{cm}^2$) one day before subculture.
3. Warm complete medium, trypsin/EDTA solution (T/E, Cat. #0103), T/E neutralization solution (TNS, Cat. #0113), and DPBS (Ca^{++} and Mg^{++} free, Cat. #0303) to **room temperature**. We do not recommend warming reagents and medium at 37°C water bath prior use.
4. Rinse the cells with DPBS.
5. Add 8 ml of DPBS and then 2 ml of T/E solution into flask (in the case of T-75 flask). Gently rock the flask to ensure complete coverage of cells by T/E solution. Incubate the flask at 37°C incubator for 1 to 2 minutes or until cells completely round up. Use microscope to monitor the change in cell morphology.
6. During incubation, prepare a 50 ml conical centrifuge tube with 5 ml of fetal bovine serum (FBS, Cat. #0500).
7. Transfer T/E solution from the flask to the 50 ml centrifuge tube (a few percent of cells may detached) and continue to incubate the flask at 37°C for another 1 to 2 minutes (no solution in the flask at this moment).
8. At the end of incubation, gently tap the side of the flask to dislodge cells from the surface. Check under microscope to make sure that all cells detach.
9. Add 5 ml of TNS solution to the flask and transfer detached cells to the 50 ml centrifuge tube. Rinse the flask with another 5 ml of TNS to collect the residual cells.
10. Examine under microscope for a successful cell harvest by looking at the number of cells being left behind. There should be less than 5%.
Note: Use ScienCell T/E solution that is optimized to minimize cell damages due to over trypsinization.
11. Centrifuge the 50 ml centrifuge tube at 1000 rpm for 5 min. Resuspend cells in culture medium.
12. Count and plate cells in a new, poly-L-lysine coated culture vessel with cell density as recommended.

Caution: Handling animal derived products is potentially biohazardous. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination [1].

[1] Grizzle WE, Polt S. (1988) "Guidelines to avoid personal contamination by infective agents in research laboratories that use human tissues." *J Tissue Cult Methods*. 11: 191-9.